Clinical and microbiological profile of post-traumatic osteomyelitis in tertiary hospital in Yaoundé – Cameroon

Handy ED, Manfo AA, Gonsu KH, Essi MJ, Ngo Nonga B and Sosso MA

DOI: https://doi.org/10.22271/ortho.2017.v3.i3p.170

Abstract

Background: Post-traumatic osteomyelitis is bone infection secondary to a trauma (open fracture or bone surgery). It is a painful and frustrating disease characterised by high rates of therapeutic failures and a costly management. However, treatment is increasingly becoming difficult because of the phenomenon of antibiotic resistance. The present study was realised to determine the epidemiological and susceptibility profile of bacterial isolates in posttraumatic osteomyelitis in a tertiary hospital in the city of Yaoundé.

Method: A descriptive cross-sectional study was carried out between November 2016 and May 2017. Patients admitted for post-traumatic osteomyelitis were recruited. After recoding clinical informations, deep specimens were taken during surgical debridement and were cultured in Blood, Mannitol Salt and MacConkey plates at 37 °C for 24-72hrs. Culture isolates were identified by standard biochemical reactions. Antibiotic susceptibility was performed by Kirby-Bauer method using the CASFM guidelines.

Results: A total of 31 patients were recruited; the modal age range was 21-50 years (67.8%) and the sex ratio 1.8:4. The commonest bones affected were the tibia (48.5%) and femur (32.3%). Predisposing factors identified included open fractures (76%), delay surgical debridement (83.3%), presence of prosthesis (58%) and surgical wound infections (23%). Out of 31 samples, 29 yielded positive culture giving rise to 48 bacteria isolates. Fourteen samples (48%) were polybacterial. The most predominant species was Escherichia coli (29%), followed by Staphylococcus aureus, Pseudomonas aeruginosa and Klebsiella pneumoniae (all 22.6%). The Gram positive organisms showed good sensitivity to Imipenem, Rifampicin, Fucidine, Lincomycin, and to Vancomycin whereas the Gram negative bacilli were mostly sensitive to Imipenen (96.7%), Amikacin (82.1%) and to a lesser extend Quinolones (54%) and Piperacillin/Tazobactam and Ceftazidime (48%).

Conclusion: Nosocomial bacteria dominate the bacterial flora of posttraumatic osteomyelitis in our setting and many multidrug resistant strains are emerging thus emphasizing on the importance of hygiene and targeted antibiotic therapy.

Keywords: Microbiological profile, Post-traumatic osteomyelitis, antimicrobial susceptibility

Introduction

Osteomyelitis can be defined as an inflammation of bony tissue caused by an infecting organism. Osteomyelitis can develop from haematogenous spread of an infectious agents, invasion of bone tissue from adjacent site of infection, or direct inoculation of germs following trauma (open fractures or bone surgery) [1]. The last entity defines post-traumatic osteomyelitis. The incidence of post-traumatic osteomyelitis is rising in our setting because of the increase in frequency of road traffic accidents and the development of orthopaedic procedures. However, osteomyelitis is a serious and invalidating illness characterised by a high rate of treatment failures and often requires long periods of treatment and hospitalisation leading to temporary impairment and at times long lasting disability or even permanent handicaps [2]. Treatment comprises of surgical debridement of all necrotic bone and soft tissue along with use of appropriate antimicrobial therapy. Treatment however is becoming increasingly troublesome due to rise in drug resistant isolates in osteomyelitis cases [3]. A good knowledge of the bacterial profile and their sensitivity pattern is therefore essential to initiate a good antibiotherapy while awaiting culture results.
**Materials and Methods**

A descriptive cross-sectional study was carried out from 15th November 2016 to 15th May 2017. Cases of post-traumatic osteomyelitis were recruited at the Orthopaedic ward of the Yaounde Central Hospital (YCH) and after obtaining patient’s consent, clinical informations were taken. The important factors taken into consideration were the patient’s age, sex, bone involved, signs, symptoms, duration of the illness and predisposing risk factors. Clinical specimens like pus, pus swabs, bone marrow contents and bone sequestrum were then collected during surgery and immediately (within an hour) sent to the Bacteriology Laboratory of Yaoundé University Teaching Hospital (YUTH) for analysis. No specific transport medium was used. The samples were cultured aerobically in Blood, Mannitol salt and MacConkey agar plates. Quality control of the media was done by verifying dates of expiration and the ensuring the absence of growth before usage. The organisms isolated were identified by routine standard operative procedures. Antimicrobial susceptibility testing was done by Kirby-Bauer’s disc diffusion method. Antibiotics tested included Ampicillin (10µg), Oxacillin (5µg), Ticarcillin (75µg), Piperacillin (30µg), Ticarcillin/clavulanic acid (75/10 µg), Piperacilin/tazobactam (30/6 µg), Amoxicillin/clavulanic acid (20/10 µg), Cefuroxime (30µg), Cefotaxime (30µg), Cefazidime (30µg), Imipenem (10µg), Vancomycin(30µg), Tobramycin (10 µg), Gentamicin (10µg), Amikacin (30µg), Ciprofloxacin (5µg), Levofloxacin (5µg), Erythromycin (5µg), Lincomycin (15 µg), Cotrimoxazole (1.25µg /23.75µg), Linezold (30µg), Rifampicin (30µg), Fluoricid acid (10µg). The results wereanalyzed using the 2016 guidelines of the « comité de l’antibiogramme de la société française de microbiologie (CASFM) ». Data obtained was entered in CSPro Version 6.3.2 software and transferred into the software SPSS Version 20 for analysis. Results were presented in counts and percentages.

**Table 1:** Different bacteria isolated

<table>
<thead>
<tr>
<th>Bacterial group</th>
<th>Species</th>
<th>Number (%)</th>
<th>Prevalence in patients (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Staphylococcaceae 7 (14.5%)</td>
<td><em>Staphylococcus aureus</em></td>
<td>7 (14.5%)</td>
<td>22.6</td>
</tr>
<tr>
<td></td>
<td>Coagulase negative</td>
<td>0 (0.0%)</td>
<td>0.0</td>
</tr>
<tr>
<td>Enterobacteriaceae 30 (62.5%)</td>
<td><em>Enterococcus sp</em></td>
<td>2(4.1%)</td>
<td>6.5</td>
</tr>
<tr>
<td></td>
<td><em>Escherichia coli</em></td>
<td>9(18.8%)</td>
<td>29.0</td>
</tr>
<tr>
<td></td>
<td><em>Klebsiella pneumoniae</em></td>
<td>7(14.5%)</td>
<td>22.6</td>
</tr>
<tr>
<td></td>
<td><em>Enterobacter sp</em></td>
<td>4(8.3%)</td>
<td>12.9</td>
</tr>
<tr>
<td></td>
<td><em>Citrobacter sp</em></td>
<td>3(6.3%)</td>
<td>9.7</td>
</tr>
<tr>
<td></td>
<td><em>Serratia sp</em></td>
<td>2(4.1%)</td>
<td>6.5</td>
</tr>
<tr>
<td></td>
<td><em>Proteus sp</em></td>
<td>2(4.1%)</td>
<td>6.5</td>
</tr>
<tr>
<td></td>
<td><em>Morganella morganii</em></td>
<td>1(2.1%)</td>
<td>3.2</td>
</tr>
<tr>
<td></td>
<td><em>Klebsiella terrigena</em></td>
<td>1(2.1%)</td>
<td>3.2</td>
</tr>
<tr>
<td>Non fermenting Gram negative bacilli 9 (18.8%)</td>
<td><em>Pseudomonas aeruginosa</em></td>
<td>7(14.5%)</td>
<td>22.5</td>
</tr>
<tr>
<td></td>
<td><em>Pseudomonas fluorescens-puiteda</em></td>
<td>2(4.1%)</td>
<td>6.5</td>
</tr>
</tbody>
</table>

All the *Staphylococcus aureus* strains were sensitive to Imipenem, Rifampicin, Fucidine, Tetracycline, Lincomycin, and to Vancomycin while 50% were resistant to Oxacillin (MRSA) (figure 1). They all showed reduced sensitivities to the other antibiotics. The two strains of *Enterococcus sp.* isolated were susceptible to most of the antibiotics except for Gentamicin, Erythromycin and Oxacillin for which only 1 strain was sensitive. The different isolates of *Pseudomonas sp.* were most sensitive to Imipenem (88.9%), Amikacin (75%) and Quinolones (66.7%). Only about one half (55.6-57%) of the isolates were sensitive to Ticarcilline-clavulanic acid and Pipercillin-tazobactam (figure 2). A few antibiotics inhibited the growth of the majority of the enterobacteria isolates. These included Imipenem (97%), Amikacin (82%) and Levofloxacin/Ciprofloxacine (54%) and to a lesser extent, Pipercillin-Tazobactam (48%), Ceftazidime (43%) and Ticarcillin-clavulanic acid (38%) (See figure 3).

**Results**

Thirty one patients with post-traumatic osteomyelitis were admitted in the orthopaedic ward within our study period and were all included in the study. We noted a predominance of the male sex with a sex ratio of 1.8:1. The age group involved majorly was between 31-40 years (29%) followed by 21-30 years (25.8%), 51-60 (19.4%), 41-50 (12.9%), and 61 years and above (12.9%). The main cause of the primary bone lesion was road traffic accidents, n= 20(65%) followed by falls, n= 7(21%) and assault, n= 4(14%). Surgery was the initial cause of bone exposure in 7 (23%) of the patients against 24 (76%) of patients who had an initial open fracture. Eighteen (58%) patients had infections on prosthetic devices. The commonest site affected was the tibia (48.5%) followed by the femur (32.3%) and the ankle (16.1%). Twenty four (83.5%) of the patients with open fractures were debrided more than 3 days after the initial injury. One (4.2%) patient was managed within the first 6 hours. 8 (26%) patients first resorted to traditional bone settlers for initial management. Sixteen patients (52%) had acute osteomyelitis (symptoms had been evolving for less than 6 weeks) while the remaining 48% had a chronic bone disease.

Out of the 31 samples analysed, 29 (93.5%) positive culture results were found; 52% being monobacterial giving a total of 48 bacterial isolates (average of 1.7 bacteria per sample). Gram negative bacteria were the most predominant isolates (81%). Enterobacteria represented 62.5% of the isolates followed by non-fermenting Gram negative bacilli (18.8%) and Gram positive cocci (18.8%). The most predominant species was *Escherichia coli* with a prevalence of 29%, followed by *Staphylococcus aureus*, *Pseudomonas aeruginosa* and *Klebsiella pneumoniae* (all 22.6%) (see Table 1).
Discussions

Clinical profile

The incidence of posttraumatic osteomyelitis was observed high among males (with a sex ratio of 1.8:1) and in the age groups between 31-40 (29%) and 21-30 (25.8%). This high predominance of the male gender and the youthful age group could be attributable to the greater likelihood of trauma and compound fractures in this population group in relation with their daily activities. These results are similar to those obtained in a study done in India in 2017 also reported a higher cases of osteomyelitis among younger age groups of 31-40 (32.8%) followed by 21-30 (25.6%) [4]. A study in Brazil, 2012 reported similar results [5].
Road traffic accidents were the major cause of primary bone injury in our study accounting for up 65% of the causes. The advent of motor cycles, the nature of roads in our setting and the non-respect of road security measures may explain these findings [6]. The next predominant cause of injury was falls (21%). Most of the cases of falls however were elderly women who are known to have reduced bone density associated with their menopausal status. These results are similar to those obtained in other studies in African context [7-9]. Of these bone lesions, 76.7% of the patients developed the bone infection after an initial open fracture. Open fractures expose the bony tissue thereby increasing the risk of inoculation of bacteria into the bone either during the accident or during wound dressing at the hospital [6]. This was even more evident in our study as almost all (96%) of the patients were debrided after 6 hours; 83% being debrided after 3 days, when local signs of infection had started appearing. Twenty six percent of the patients resorted to traditional bone settlers for initial management. A retrospective study done in Mali from 1994-2003 reported that 57% of the patients first consulted traditional doctors before consulting at hospitals [10]. This constant resort to traditional bone settler could be explained by ignorance and poverty in our setting. Post-surgical wounds were the predisposing factor in 23.3% of the patients in our study. These were probably a consequence of defects in the respect of hygienic conditions during surgery or wound dressing. Metal works (implants) were associated with 58% of infections. In our study, predominant involvement of the lower limb was noticed with the tibia, the femur and bimalleolar accounting for 48.5%, 32.3% and 16.1% respectively. The tibia because of its anatomic situation (proximity to the skin) is the bone that is most susceptible to open fractures and therefore to osteomyelitis.

In a study realized in India in 2013 [11], the major predisposing factors identified included open fractures (53%), post-operative infections (26%) and implants (19%). The most affected bones were the same as for our study and included the tibia (58%) and the femur (31%). Another study realized in India in 2016 [12] identified as predisposing factors open fractures (45.6%), post-operative infections (27.2%) and implants (24.0%). The most affected bones also included tibia and the femur (44.0% and 40.8% respectively). The higher percentage of open fractures in our study could be explained by the fact that we only considered post traumatic osteomyelitis meanwhile those studies also included haematogenous osteomyelitis and osteomyelitis on diabetic foot ulcers. With respect to duration of illness, 52% of our patients had an acute infection while 48% had a chronic infection. In the study done in India in 2016, 77.8% of the patients had chronic osteomyelitis in contrast to 22.2% who had an acute infection. However, they included in their study haematogenous osteomyelitis and diabetic foot ulcers in their study.

Culture results
In our samples, 93.5% gave positive culture results and enabled the isolation of a total of 48 isolates giving an average of 1.4 bacteria per sample. Overall, 52% of the samples were monobacterial while 48% were polybacterial. Our results are similar to those reported in a similar study done in Morocco, in 2007, which reported the average of 1.4 bacteria per specimen and 63.5% were monomicrobial against 36.5% of the cases which were polymicrobial [12]. The study realized in India in 2010 reported 72% of positive cultures, 70% of these being monobacterial cultures and 30% polybacterial [13]. In our study, Gram negative bacilli were the most common bacteria isolated (62.5%) with Escherichia coli being the most predominant organism (18.8%) followed by Staphylococcus aureus, Pseudomonas aeruginosa and Klebsiella pneumonia (14.5%). This corresponds to the profile of nosocomial germs showing the latter a major role in posttraumatic osteomyelitis in our context. This does not correlate with most studies as Staphylococcus aureus is often reported to be the first causative agent [4, 11, 14, 15]. A similar study done in Iraq in 1998 however reported Pseudomas sp, Klebsiella sp and Proteus sp to be more predominant than Staphylococcus aureus [16].

Sensitivity testing
All the strains of Staphylococcus aureus were sensitive to Imipenem, Rifampicin, Tetracycline, Fludic acid, Lincomycin and to Vancomycin (VSSA) while 50% were susceptible to Oxacillin (MSSA). Their sensitivity for quinolones, aminoglycosides, Cotrimoxazole and betalactams (including combination forms with betalactamase inhibitors like Amoxicillin-Clavulanic acid) was generally low (less than 50%). These findings are different from those obtained in the study carried out in Morocco 2014 [12] which reported a good (87-100%) sensitivity of all the Staphylococcus aureus strains to most antibiotics except for Penicillin G. All the strains were methicillin and Vancomycin sensitive. The difference could be explained by the fact that they mostly had community acquired germs. The studies carried in India in 2013 [11] and 2017 [14] had comparable results with ours. They showed reduced sensitivity (less than 50%) to quinolones, aminoglycosides, Cotrimoxazole and to Cephalosporins. The percentages of MSSA were 56.9% and 76.1% respectively and all samples in both studies were however sensitive to Vancomycin. The overall sensitivity of the strains of Enterococcus sp. were generally high correlating with the findings in a similar study done in India in 2016 [15]. The isolates of Pseudomonas sp. were most sensitive to Imipenem (88.9%), Amikacin (75%) and Quinolones (66.7%) followed by Ticarcilline-clavulanic acid and Piperacillin-tazobactam (55.6 and 57% respectively). They showed reduced (12.5-25%) sensitivites to the other aminoglycosides, Amoxicillin-clavulanic acid and cephahlosporins. The study in India 2013 [11], reported a best sensitivity rates to be amongs Piperracin-Tazobactam (89%), Amikacin (78%) and the Quinolones (67-89%). The sensitivity to cephahlosporins was also low but that to aminoglycosides relatively higher (67%). These findings also correlate to the 2003 report of “The Surveillance Network, TSN “USA [17]. The study in Morocco 2014 reported a good (more than 50%) sensitivity to most antibiotics except for Amoxicillin-Clavulanic acid, Ampicillin and Cefalotin [12].

Most of the Enterobacteria in our study were sensitive to Imipenem (97%), Amikancin (82%) and quinolones (54%) and about 50% were either sensitive or intermediate to Piperracin/Tazobactam, and Ceftazidime. They showed high resistance to aminoglycosides, other cephahlosporins, and Cotrimoxazole. This susceptibility rate is relatively low compared to the findings of “The Surveillance Network, TSN “USA which reports high sensitivities (>78%) of the different enterobacteria strains to quinolones, third generation cephahlosporins, and piperacillin/Tazobactam [17]. This difference could be explained by the fact that most of these organisms in our study were probably hospital acquired. The findings however correlate with those of the study done in India in 2013 [11] where a higher sensitivity among
enterobacteria was noted only to Imipenem, Amikacin and Quinolones. Another study done in India, 2014 on chronic osteomyelitis reported even lower rates sensitivities (less than 50%) for most antibiotics except Imipenem and Amikacin. However, while antimicrobial therapy is desirable in the control of osteomyelitis, the most important factor for a successful treatment of patients with bone infection is the quality of debridement. The debridement should be done early enough and must achieve a clean and viable wound. Appropriate therapy of posttraumatic osteomyelitis includes adequate drainage, thorough debridement, obliteration of dead space, stabilization when necessary, wound protection, and specific antimicrobial therapy.

**Conclusions**

Nosocomial bacteria dominate the bacterial flora of posttraumatic osteomyelitis in our setting and these bacteria are developing resistances to most of the routinely used antibiotics. Appropriate implementation of hygienic practices and appropriate selection of antibiotics based on cultures results will help treat the disease successfully in the early stages and prevent the spread of multidrug-resistant strains to limit morbidity.

**Acknowledgement**

None

**Ethical considerations**

The manuscript has received approval from the ethical committee of the Faculty of Medicine and biomedical sciences of the University of Yaoundé I, Cameroon

**References**